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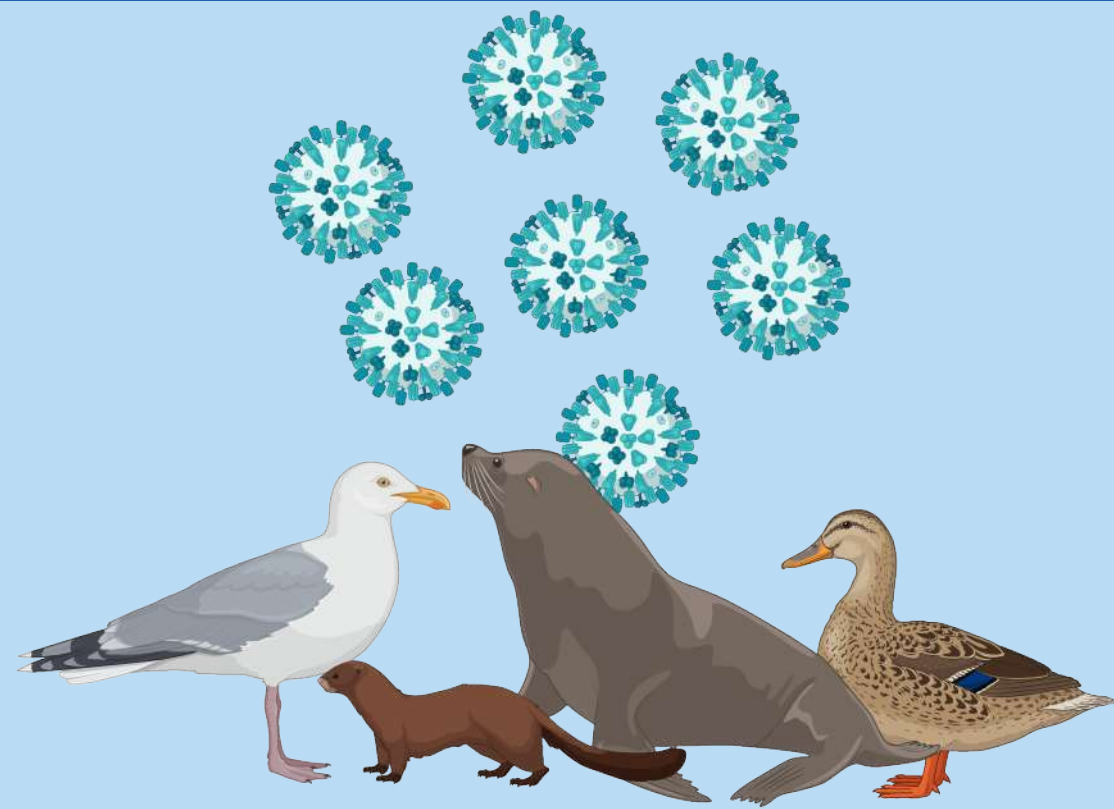
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## Non-invasive real-time monitoring



Avian influenza virus (AIV) is an RNA virus that can affect wild animals, livestock, and humans. Wild aquatic birds are the main reservoir of AIV and spread the virus around the globe, causing devastating outbreaks in wild and domestic populations. Early in situ detection and sequencing of AIV in wild populations is therefore essential to control transmission during an outbreak, understand global transmission patterns, and avoid devastating impacts on biodiversity, global health, and food security. Here, we present a pilot study to test direct RNA sequencing for developing a non-invasive real-time approach to rapidly and efficiently detect and sequence AIV through the East Atlantic flyway.



**Collection of aerosol and faecal samples**

**Field-friendly RNA extraction**

M1 Sample Prep® Cartridge Kit For RNA 2.0

**Nanopore sequencing & bioinformatic analysis**

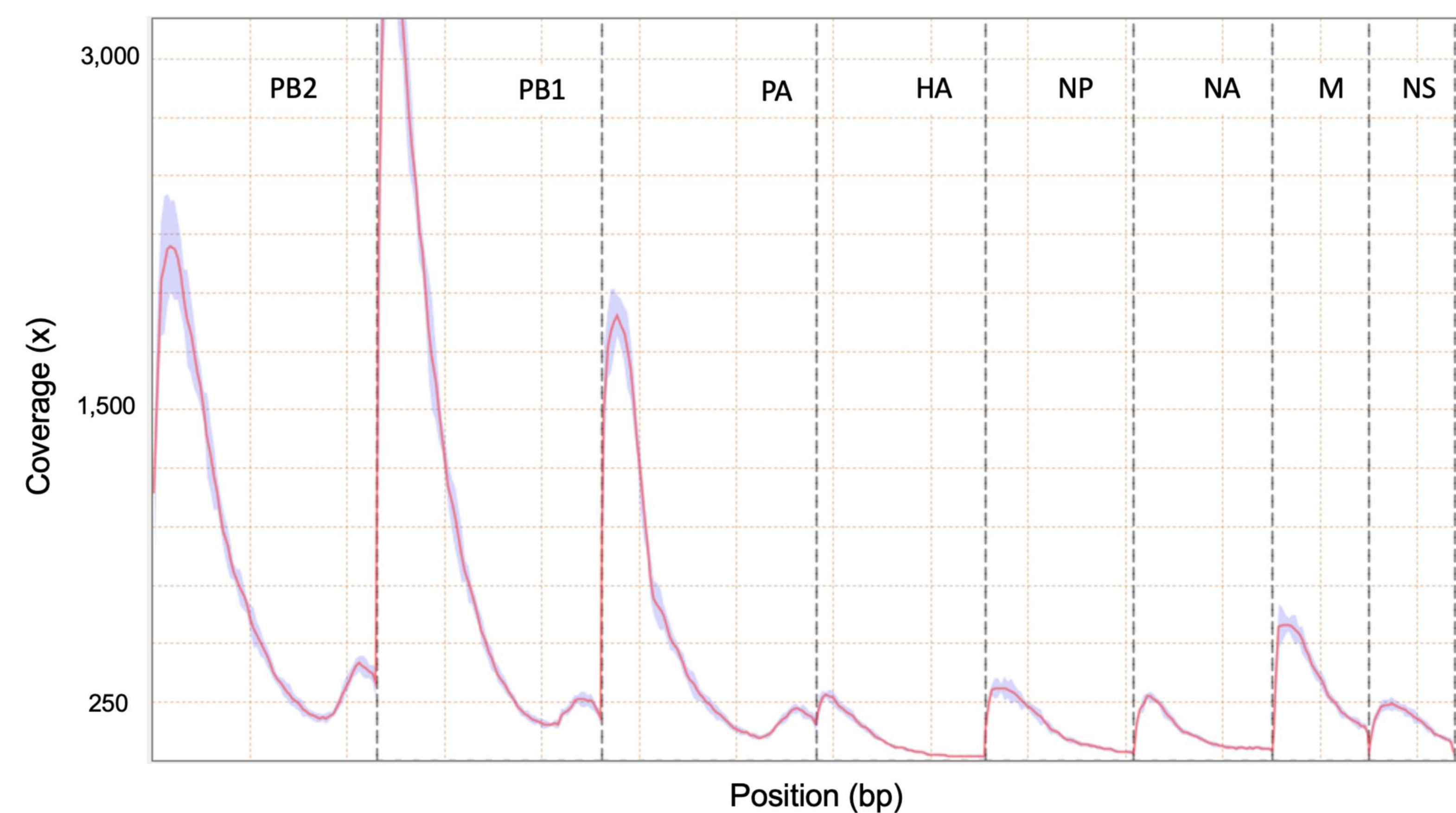
MinION

## Direct RNA sequencing

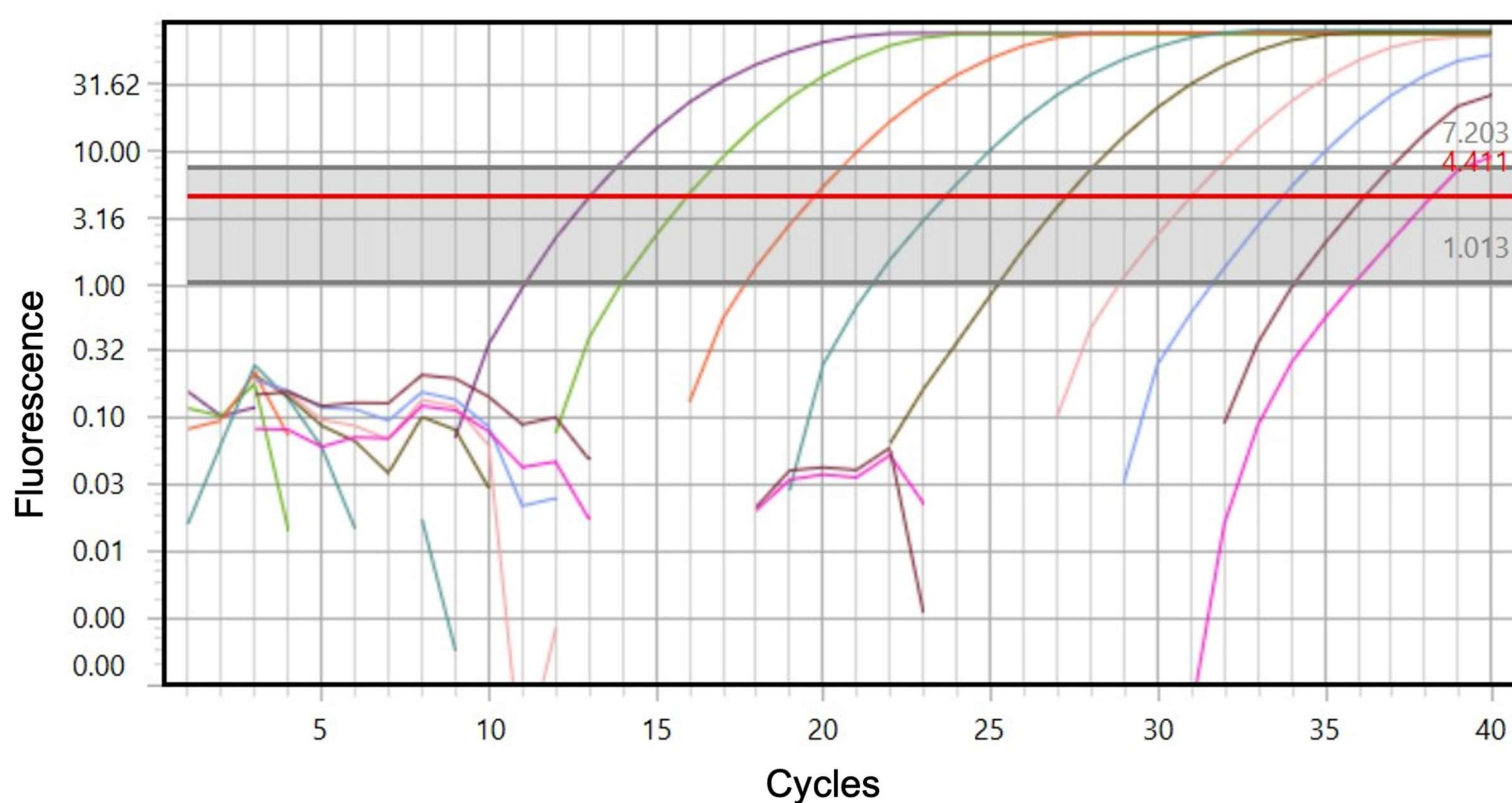
### METHODS

- 1 **Collection of virus from eggs**
- 2 **RNA extraction**  
  
NucleoSpin RNA Virus
- 3 **Viral quantification**  
  
Portable Mic qPCR Cyclers
- 4 **RNA library preparation**  
  
Adapted RTA
- 5 **Direct RNA sequencing**  
  
Direct RNA kit MinION
- 6 **Bioinformatic analysis**  
  
Guppy + minimap2

### RESULTS NATIVE VIRAL RNA SEQUENCING



### RESULTS qPCR



Cycle thresholds (Ct) of an avian H1N1 serial dilution were analyzed with a rRT-PCR targeting the M segment of the virus using a portable Mic qPCR cyclers. The original sample, with a Ct of 13, was direct RNA sequenced with the M1k.

### Influenza A virus (A/duck/Italy/281904/2006(H1N1))

|                  |        |
|------------------|--------|
| <b>Reads</b>     | 15,454 |
| <b>Mapped</b>    | 10,029 |
| <b>% Mapped</b>  | 64.9%  |
| <b>Consensus</b> | 99.61% |

| Name | Length | Mapped bases | Mean coverage |
|------|--------|--------------|---------------|
| PB2  | 2308   | 1876669      | 813.1148      |
| PB1  | 2310   | 2413382      | 1,044.7541    |
| PA   | 2200   | 1225865      | 557.2114      |
| HA   | 1741   | 166651       | 95.7214       |
| NP   | 1530   | 223824       | 146.2902      |
| NA   | 1424   | 168240       | 118.1461      |
| M    | 989    | 342473       | 346.2821      |
| NS   | 890    | 148519       | 166.8753      |

## CONCLUSION

With direct RNA sequencing, we have been able to obtain good results (threshold of >10x coverage and >90% consensus identity). For our sample with low Ct, we have even been able to achieve >100x coverage and >99% consensus identity using kit 9 chemistry. This approach, together with the detection of the virus by rRT-PCR with portable Mic PCR, is a straightforward methodology for avian influenza surveillance and sequencing in the field. However, we expect to obtain significantly lower Ct values from environmental samples, so we will next test the sensitivity of direct RNA sequencing for our diluted samples. While cDNA sequencing after multi-segment amplification is likely to be more sensitive for such samples, we will test if the new nanopore chemistry can further improve direct RNA sequencing results.

## REFERENCES

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