

# Analysis of the Elements Involved in the Enrichment of a Panel of Genomic Regions by Nanopore Sequencing Using Adaptive Sampling

Llamas-López M<sup>1</sup>, Garrido-Rodríguez P<sup>1</sup>, Cuenca-Guardiola J<sup>2</sup>, Navarro Manzano E<sup>1</sup>, Padilla Ruiz J<sup>1</sup>, Ayala F<sup>1</sup>, Macías JA<sup>1</sup>, de la Morena Barrio ME<sup>1</sup>, Fernández-Breis JT<sup>2</sup>, Lozano ML<sup>1</sup>, de la Morena Barrio B<sup>1</sup>, Corral J<sup>1</sup>

<sup>1</sup> Servicio de Hematología, Hospital Universitario Morales Meseguer, Centro Regional de Hemodonación, Universidad de Murcia, IMIB Pascual Parrilla, CIBERER-ISCIII, Murcia, Spain  
<sup>2</sup> Departamento de Informática y Sistemas, Universidad de Murcia, IMIB Pascual Parrilla, Murcia, Spain

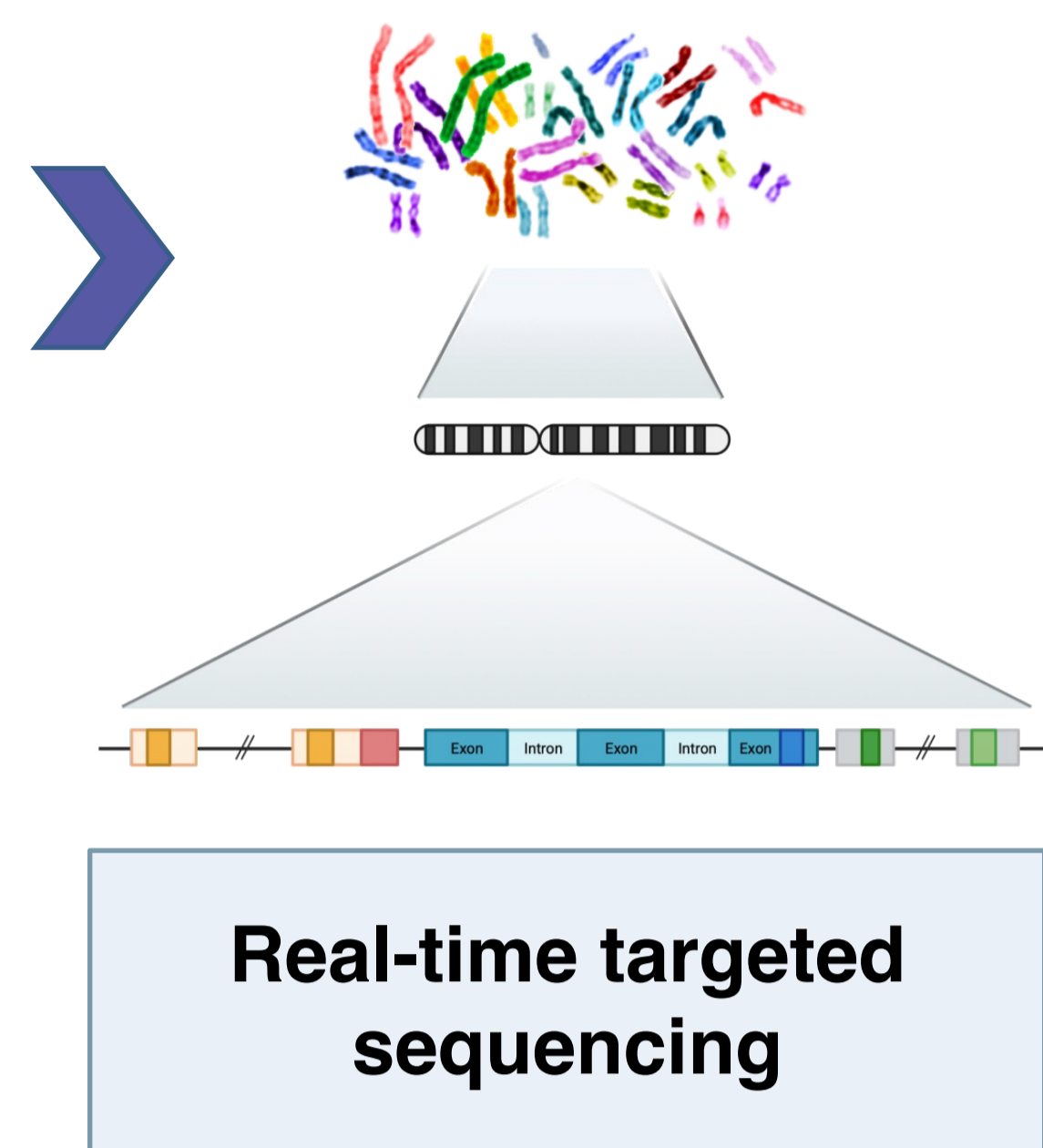
\*Oxford Nanopore Technologies products are not intended for use for health assessment or to diagnose, treat, mitigate, cure, or prevent any disease or condition.

LC

## Background

One of the main limitations of nanopore sequencing is the **low coverage** obtained.

Different methods may increase coverage, but **adaptive sampling** is the only one which does not require manipulation and can be used in MinION.

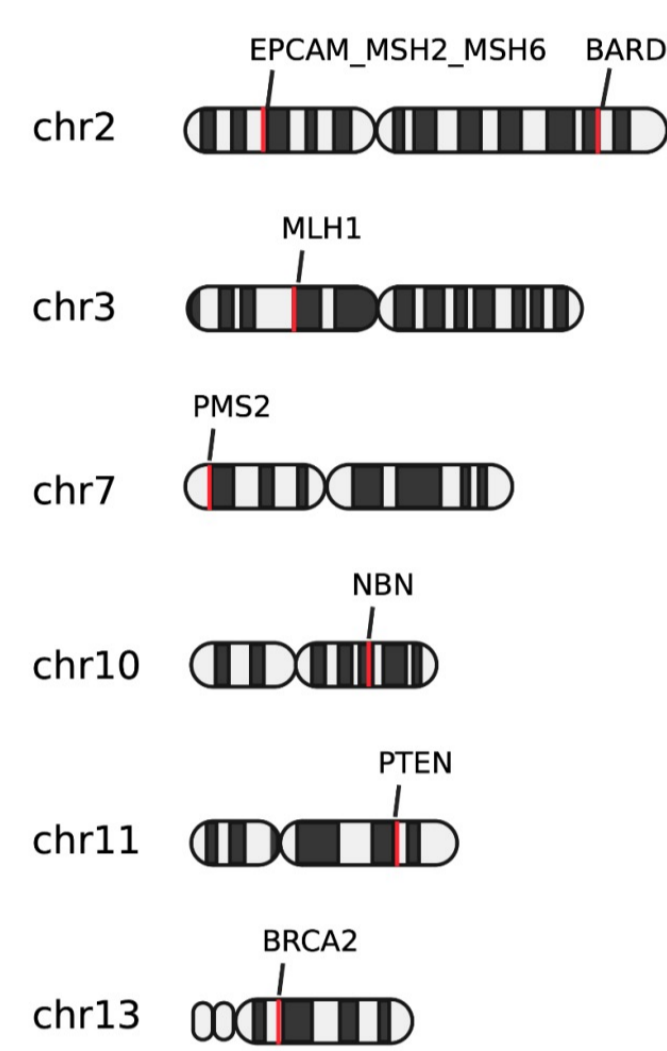


**AIM:** To dissect the factors involved in the **enrichment** process using adaptive sampling

## Materials & Methods

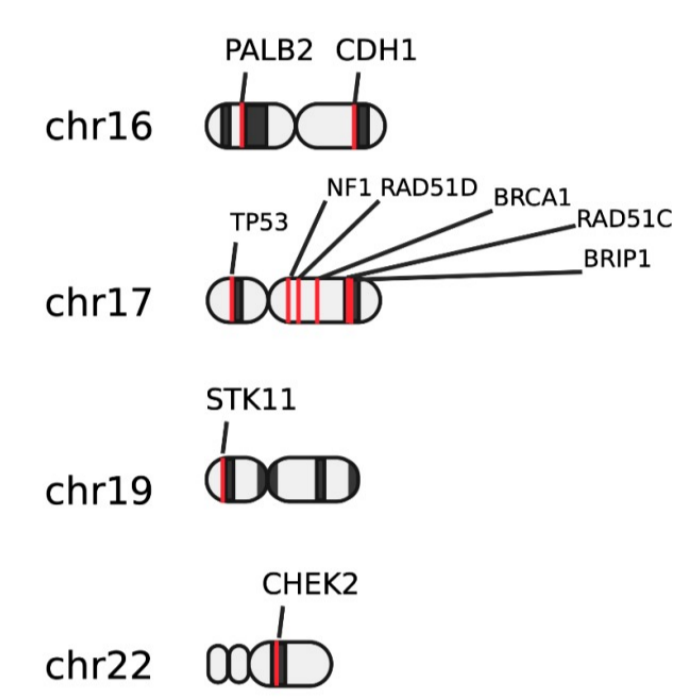
### Samples

**Sequencing: 18 regions involving 20 genes**



Negative for NGS screening

**16 cases with familial breast cancer**



Region length: **126 Kb to 565 Kb**

### Nanopore sequencing



**8** Length selection (>1,000 kb)

**5** Without length selection

Samples sequenced **twice** with **two different** settings:

**3** With and without length selection

**1** DNA fragmentation (g-TUBE Covaris® for 6 Kb fragments)

**Different sequencing times**

## Results

**Coverage reached with this method:**

**Region of Interest**  
5.1x

Up to **13.0x**

**Rest of the Genome**  
2.0x

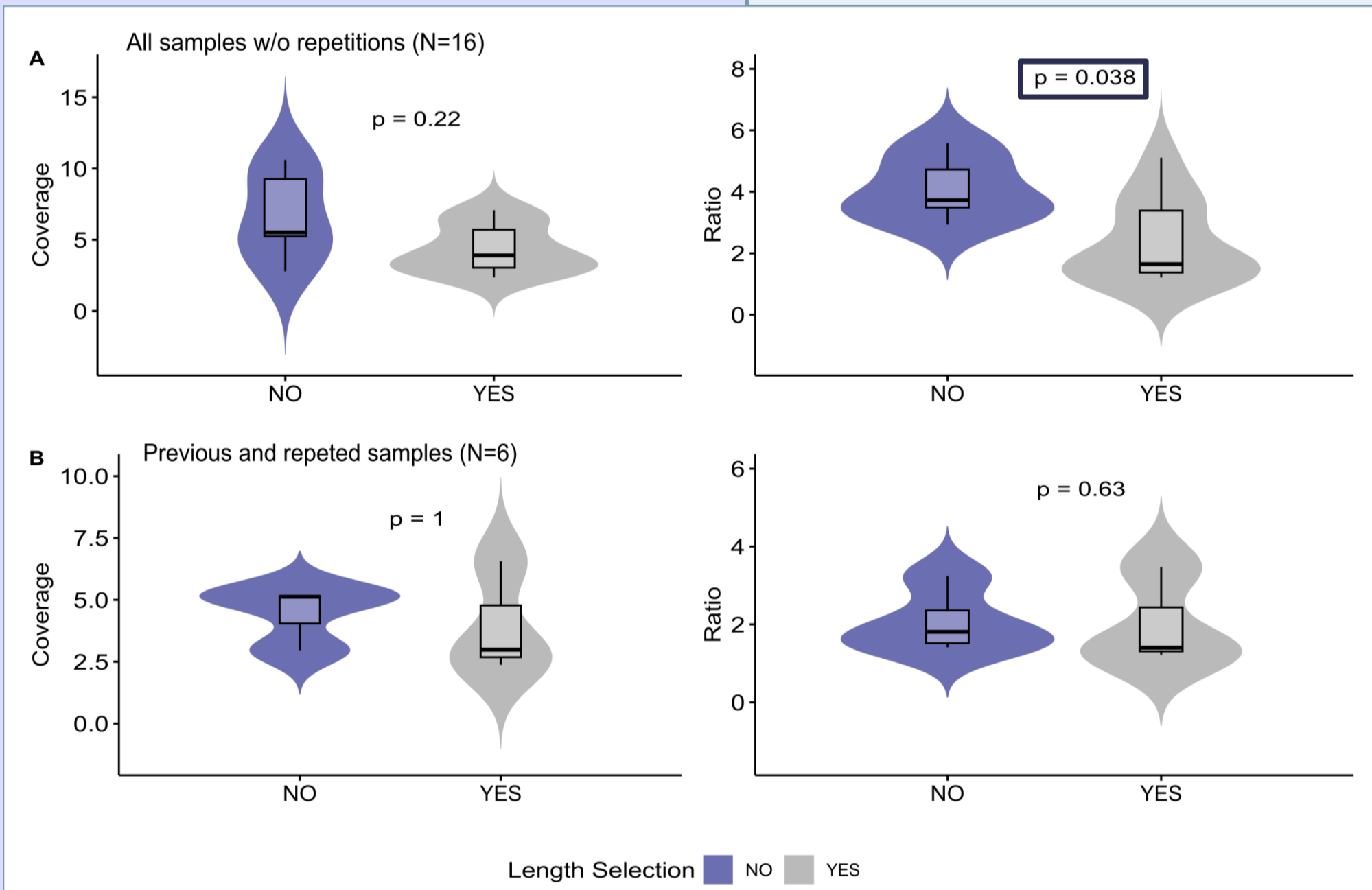
Up to **3.2x**

We have analyzed the **parameters** and **elements** potentially involved in the enrichment process by using two different approaches:

### 1) Analysis of samples:

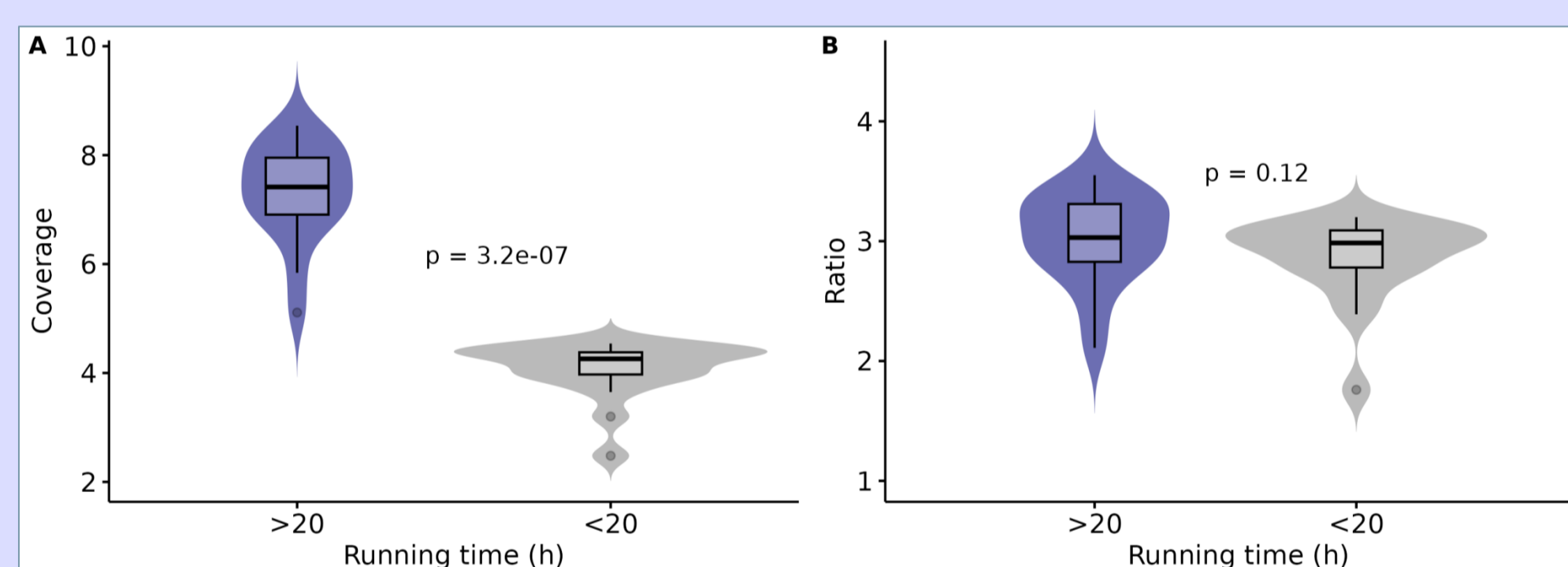
#### Long Read Selection

$$\text{Ratio} = \frac{\text{Coverage area}}{\text{Coverage genome}}$$



**Figure 1.** Average coverage and enrichment (ratio) with or without long read selection. **A)** Full cohort of samples **B)** Three Paired samples

#### Sequencing time



**Figure 3.** Average coverage **A)** and enrichment **B)** as a function of the running time

**Sequencing time (>20 h)** is the main element improving the coverage of selected regions. Likewise, it also improves coverage in the rest of the genome.

**Selection of long reads (>1,000 Kb)** and **DNA fragmentation** did not improve the enrichment yield.

### 2) Analysis of regions:

One region was **less covered**: **PMS2**.

#### Elements evaluated

Repetitive elements

Coding regions

Pseudogenes

Homopolymers

Did not affect the process

	<b>PMS2</b>	Rest of the regions
<b>Average</b>	<b>3.3x</b>	<b>4.5x</b>

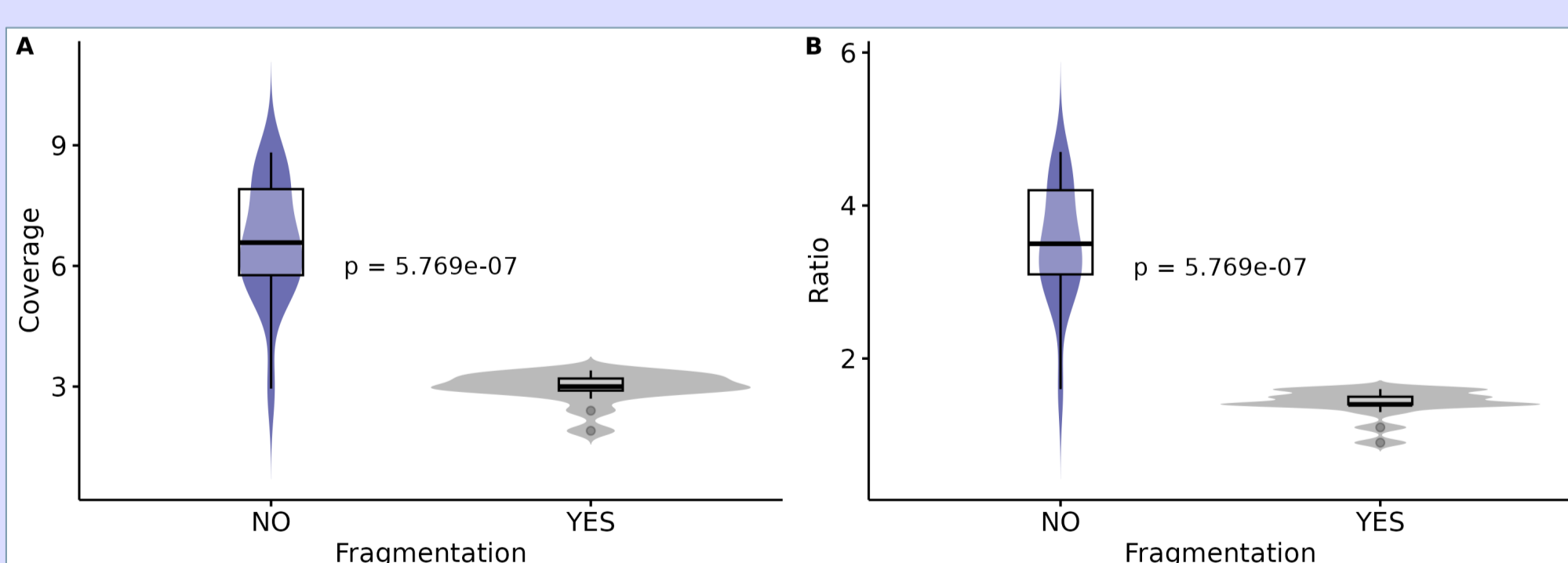
**Table 1.** Regions with a high number of pseudogenes

	<b>NF1</b>	<b>CHEK2</b>	<b>PMS2</b>
	12	7	14

**Coverage of other pseudogenes:**  
Similar or higher than the gene of interest

**High homology between genes and pseudogenes**

#### Fragmentation



**Figure 2.** Average coverage **A)** and enrichment **B)** of regions of interest according to DNA fragmentation

## Conclusions

Analysis of factors that could influence the effectiveness of adaptive sampling in a panel of 18 genomic regions using the accessible MinION device revealed these **two factors** as the **most relevant** for providing **higher coverage** in the areas of interest:

1

**Sequencing time**

2

**Presence of pseudogenes**