



Assessing the microbiome diversity and antibiotic resistance genes of African and Asian *Diaphorina citri* populations using Oxford Nanopore sequencing

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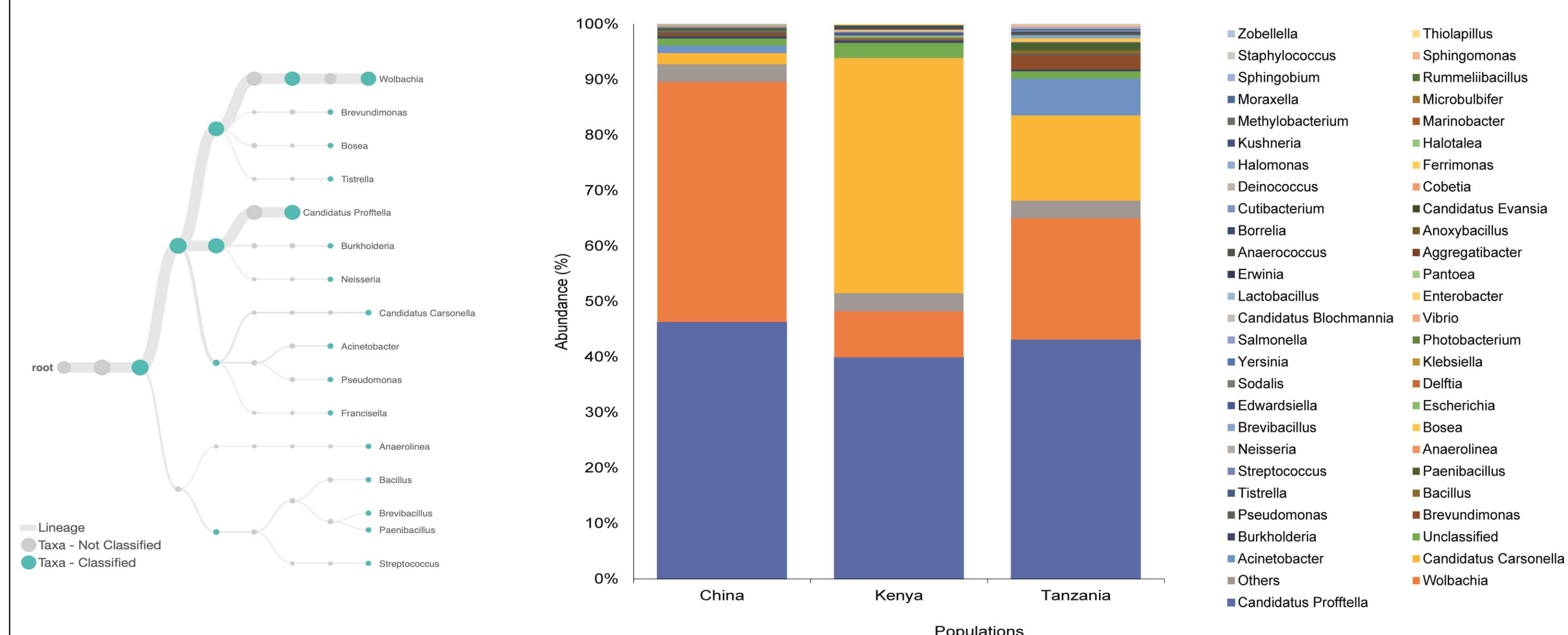
INTRODUCTION

- Diaphorina citri* is a key pest of *Citrus* sp. Worldwide, as it acts as a vector for *Candidatus Liberibacter asiaticus* (Las), the bacterial pathogen which causes the destructive Huanglongbing (HLB) disease.
- The study of microbial communities through metagenomics helps to understand their interactions with arthropods, disease transmission, immunity, host resistance and insect host vector competence.
- Recent reports of the presence of *D. citri* in Tanzania and Kenya [1,2] and Las in Ethiopia and Kenya [3,4] highlights the need to profile the microbiota diversity in the insect vector populations.
- The identification of key endosymbionts and other bacteria, as well as antibiotic resistance genes in the microbial community within the insect can inform the diversity between the newly introduced African populations and older populations.



RESULTS

1. Taxonomic assignment and Cumulative abundance of bacterial microbiota in *D. citri*



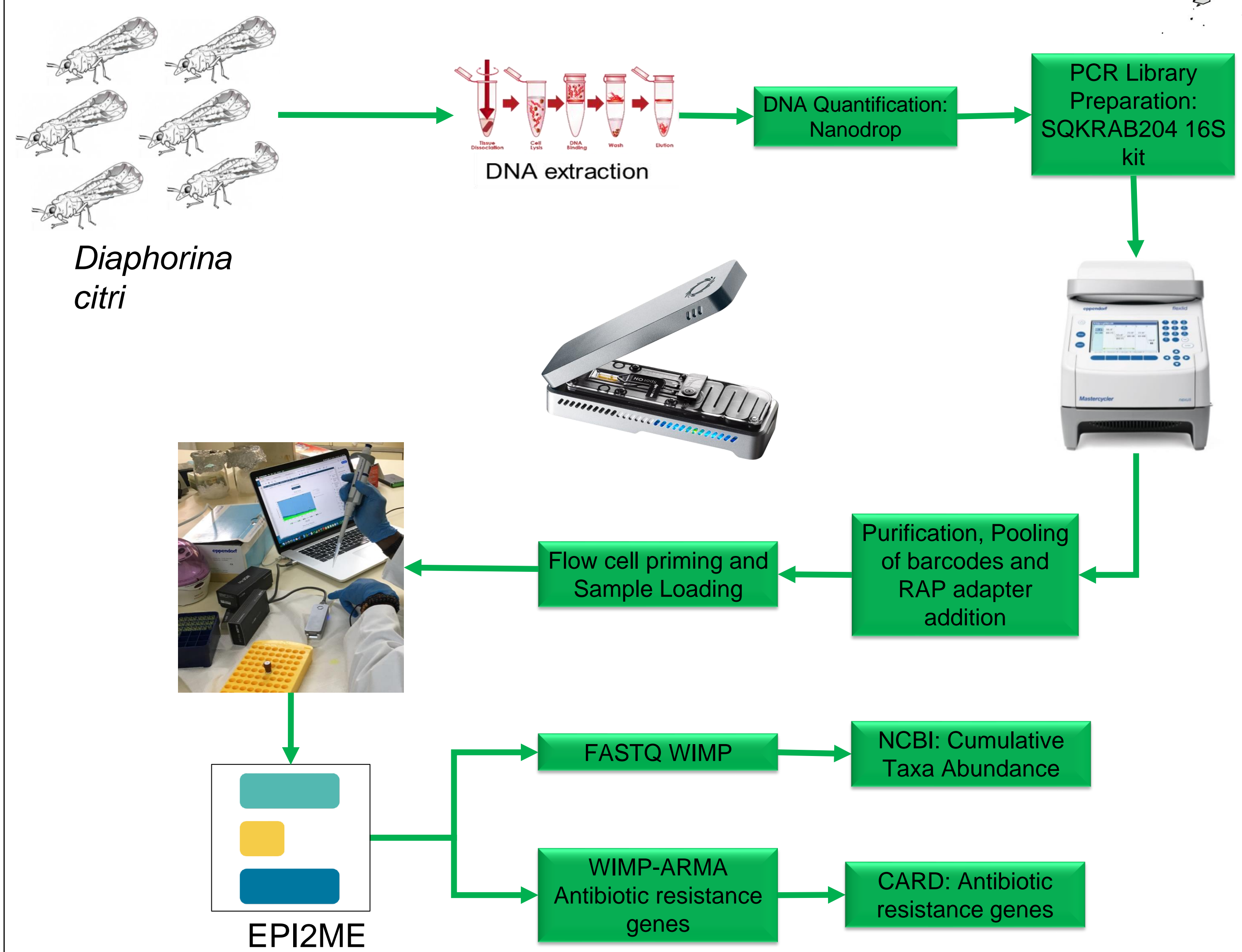
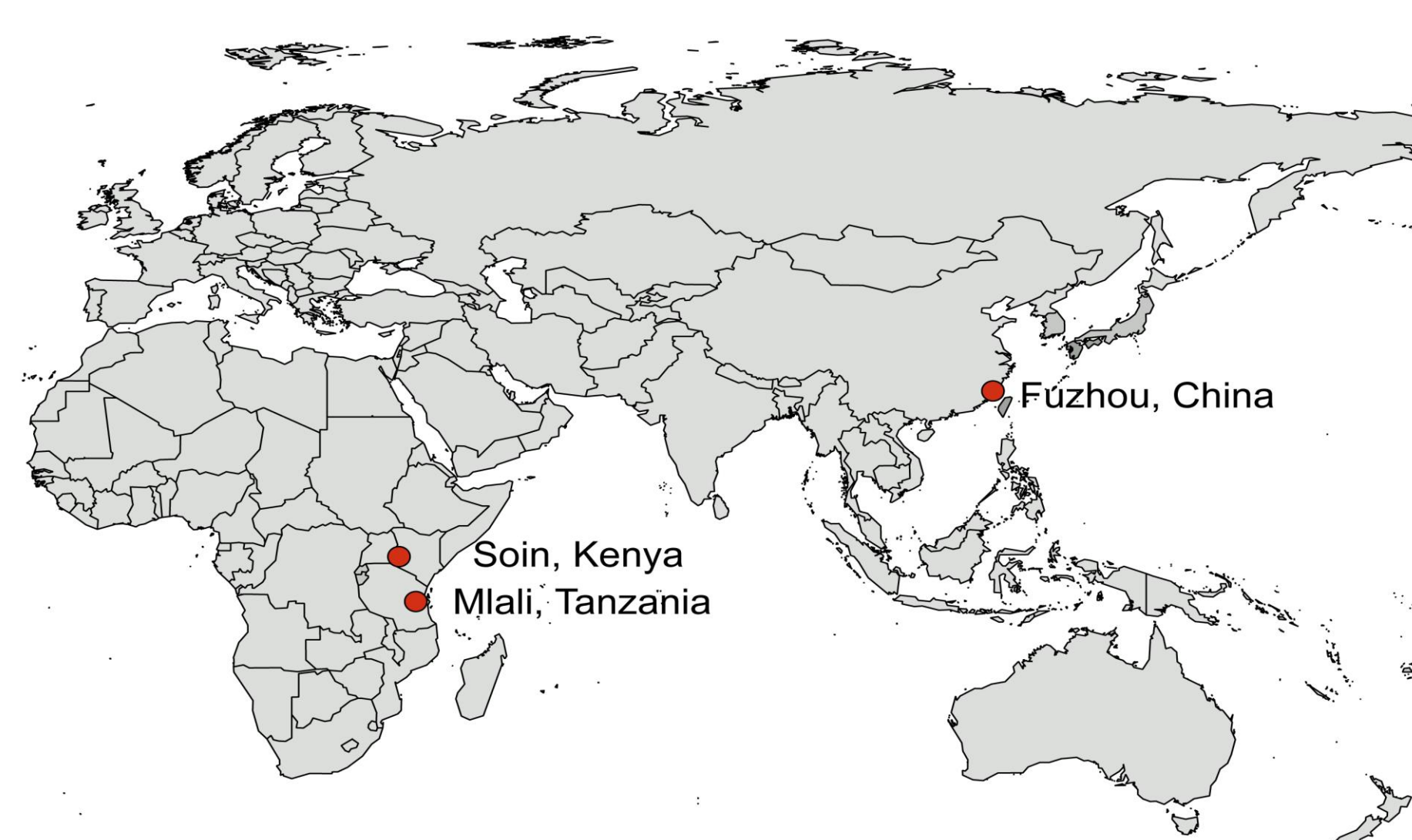
Aim

- This study sought to evaluate the bacterial diversity and genes for resistance to antibiotics within the microbiome of *D. citri* populations.

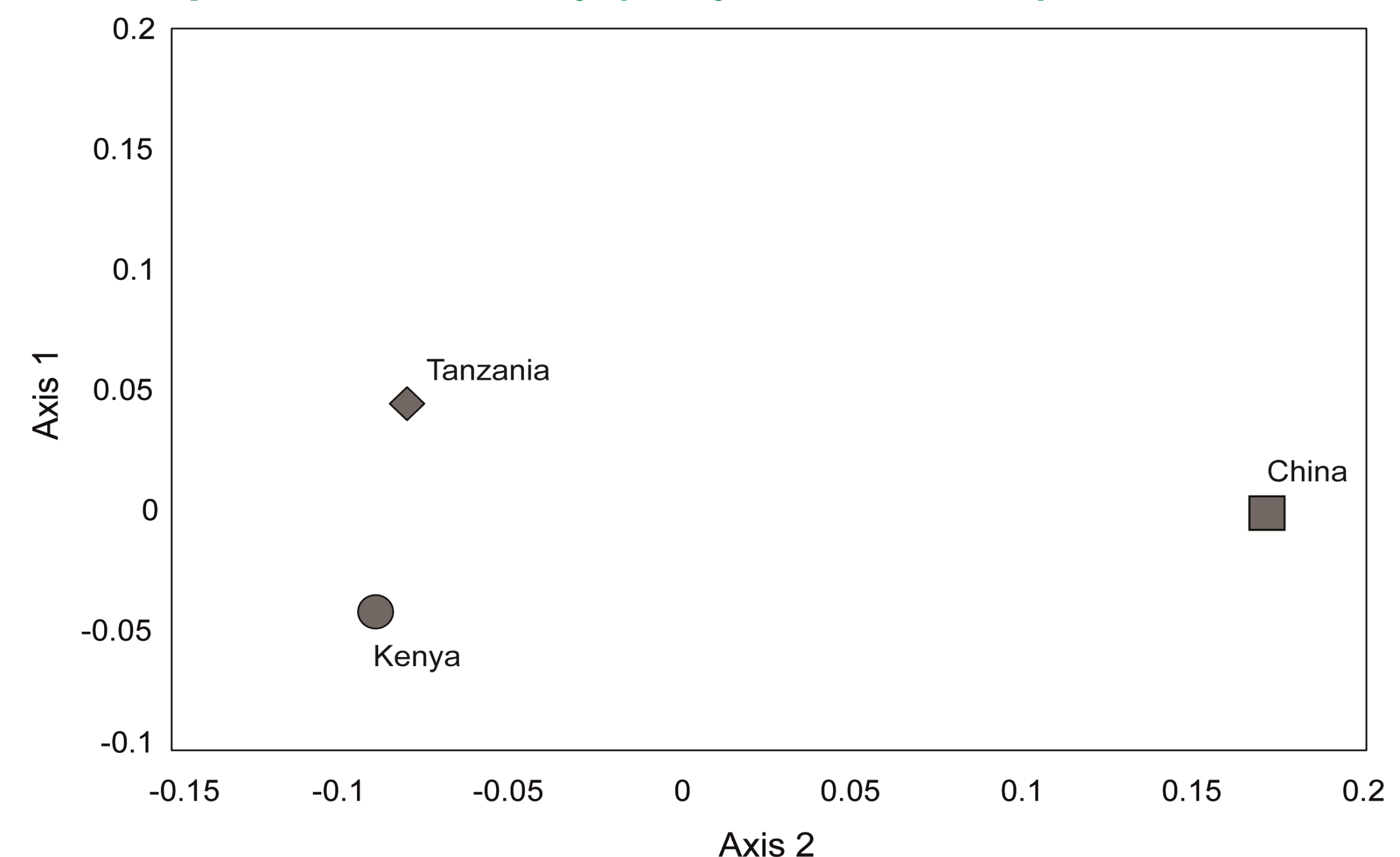
METHODS

Sampling Locations

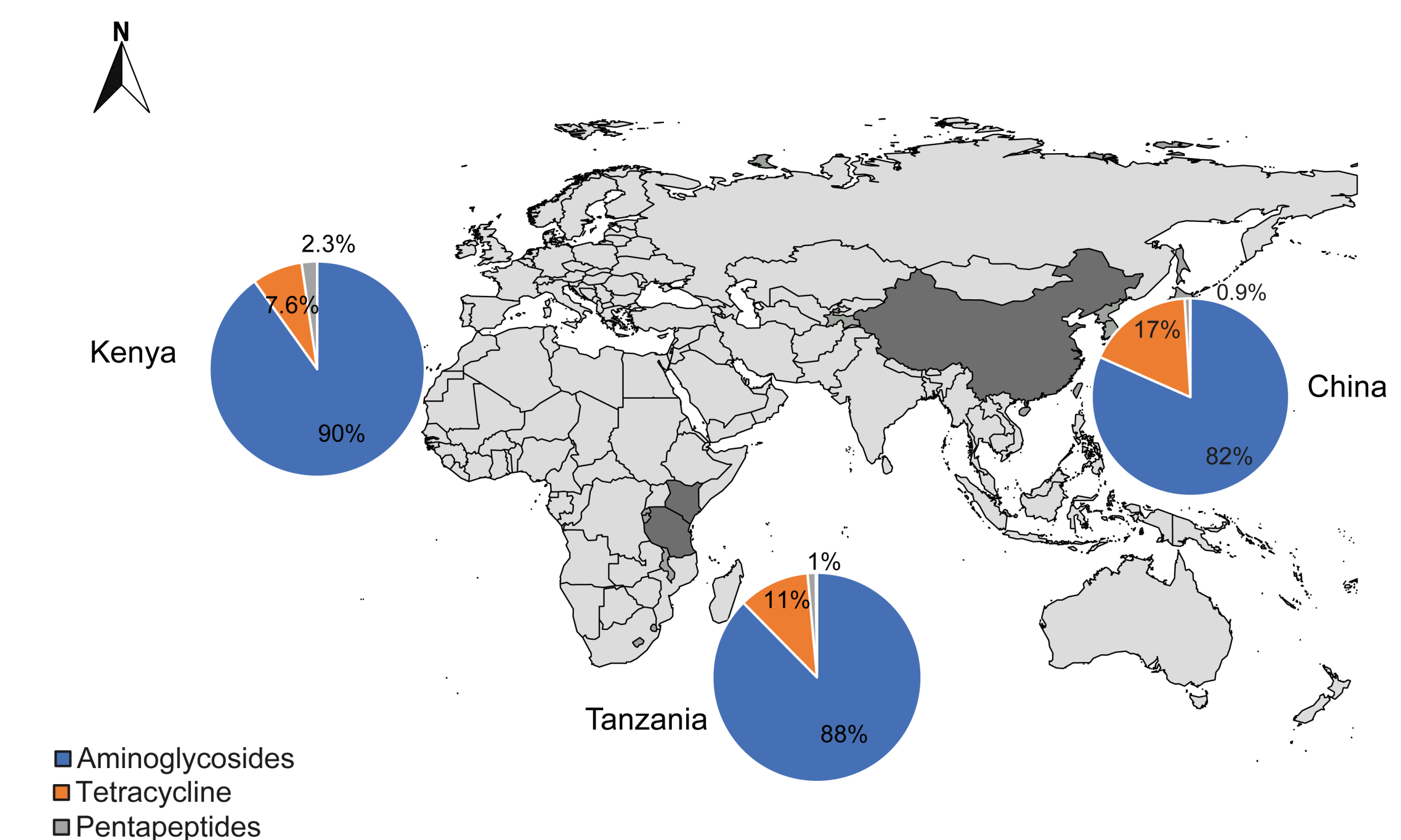
- Fuzhou, China
- Soin, Kenya
- Mlali, Tanzania



2. Inter-Population Diversity (Bray-Curtis index)



3. Abundance of genes for resistance to antibiotic compounds



CONCLUSIONS

- The most abundant genera in all populations were the Key endosymbionts of *D. citri* ("Candidatus Proffella", *Wolbachia* and "Candidatus Carsonella").
- The *D. citri* from Tanzania had the highest bacterial diversity, while Kenya and China were the most diverged populations.
- The most abundant genes for resistance to conferred resistance to aminoglycosides, tetracycline and pentapeptides families of antibiotics.

IMPACT

- The elucidation of the bacterial microbiota and its ARGs in *D. citri* provide valuable information on potential integrated pest control measures against the psyllid and its ability to transmit diseases. The Exploration of cytoplasmic incompatibility conferred by *Wolbachia* can be used to interrupt the capacity of the insect to transmit diseases as well as the influence of *Wolbachia* on the increased susceptibility of the psyllid to parasitoids.

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Acknowledgment: We gratefully acknowledge the financial and technical support of our core donors: Swiss Agency for Development and Cooperation (SDC), Switzerland; Swedish International Development Cooperation Agency (Sida), Sweden; UK Aid, Government of the United Kingdom; Ministry of Higher Education, Science and Technology, Kenya; and Government of the Federal Democratic Republic of Ethiopia. *The views expressed herein do not necessarily reflect the opinion of the donors.*



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