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Novel screening platform for
highly multiplexed biomarker analysis

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ABSTRACT | To date, most diagnostic tests rely upon the detection of a limited number of biomarkers. This often results in a lack of specificity to provide enough clinically useful detail on the pathology. To overcome this challenge, we developed a new strategy, which combines nanopore sequencing^{1,2} with barcoded nucleic acid probes, engineered to recognise a panel of biological targets (miRNAs, proteins, and small molecules). Our workflow is rapid, from sample preparation to results in 1 hour. The established method is easily adaptable, as the number and type of targets can be greatly expanded. Preliminary results indicate a significant advantage over existing techniques, which makes us believe that this approach could have an extensive impact on diagnostics and facilitate the transition to personalised medicine³.

CONCEPT

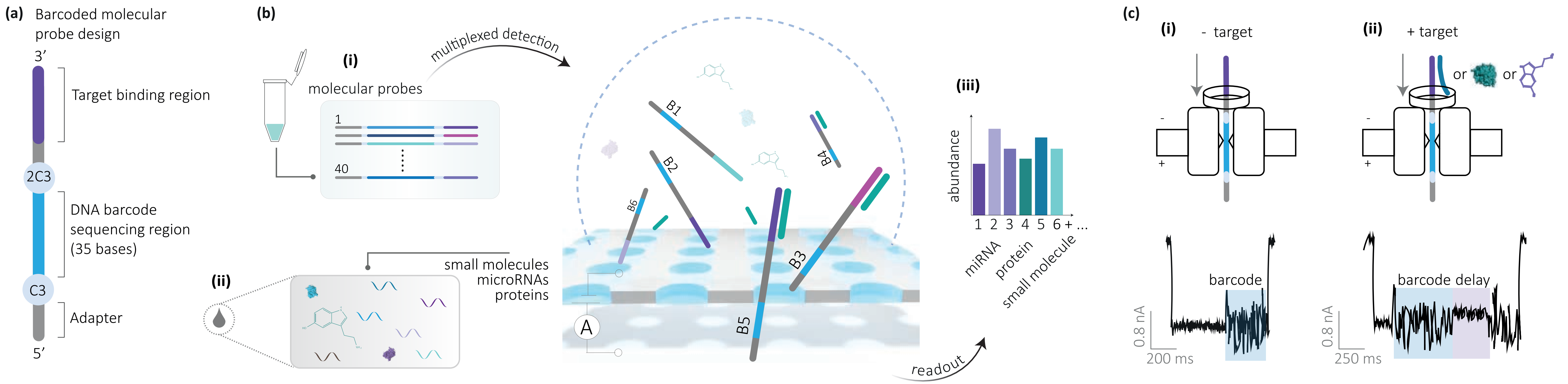


Fig. 1 Concept of multiplexed biomarker detection. (a) **Barcoded molecular probe design**⁴. The barcoded molecular probe consists of mainly three parts: 1) a Y-adapter (ONT), 2) the DNA barcode sequencing region (also called barcode), and 3) the target binding region (complementary miRNA sequence or aptamer). (b) **Workflow**. (i) Barcoded molecular probes were incubated with synthetic targets or (ii) serum samples. Nanopore sequencing of the barcode was performed to classify the barcoded molecular probe, and (iii) the presence of targets was determined by delay analysis. (c) **Delay analysis**. (i) **Without target**. Analysis of events without target result in current traces without delay. (ii) **With target**. Analysis of events with analyte-bound result in current traces with delay, due to the unzipping of the double-strand or unraveling of the aptamer.

MULTIPLEXED DETECTION OF miRNAs, PROTEINS, AND SMALL MOLECULES

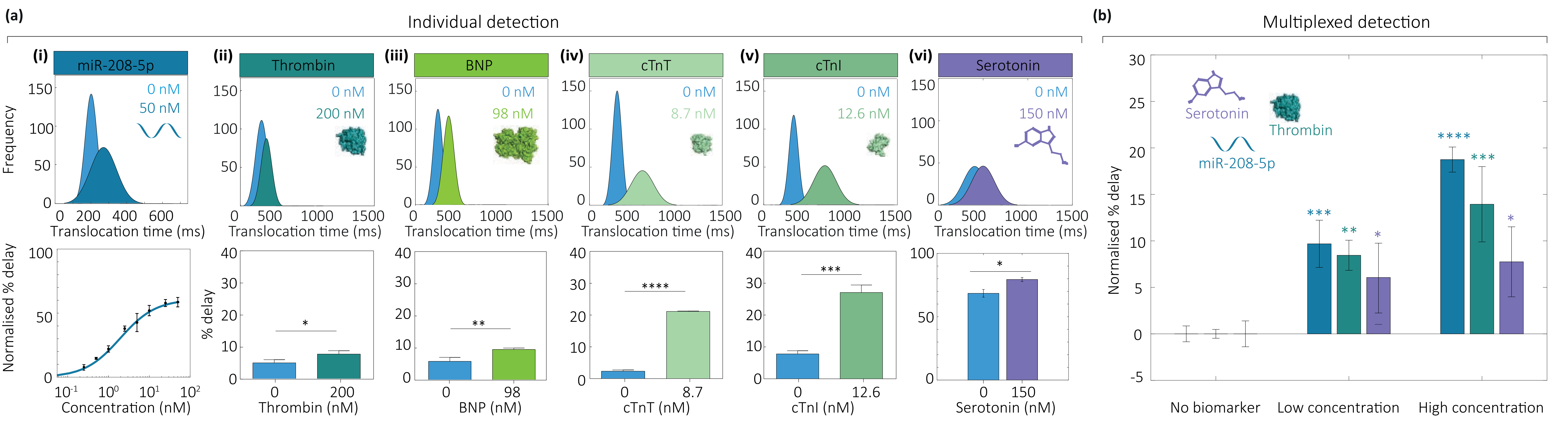


Fig. 2 Demultiplexing of miRNAs, proteins, and small molecules. (a) **Individual detection of biomarkers**. Specific barcoded molecular probes (30 nM) +/- corresponding (i) miR-208-5p, (ii) Thrombin, (iii) BNP, (iv) cTnT, (v) cTnI, and (vi) serotonin. Significance was determined by Student's t-test (*p<0.05, **p<0.01, ***p<0.001, ****p<0.0001, N=3, n=1024-3395). (b) **Multiplexed detection of biomarkers**. Low concentrations (10 nM miR-208-5p, 150 nM serotonin, 300 nM thrombin) and high concentration (50 nM miR-208-5p, 750 nM serotonin, 1500 nM thrombin) were compared to the normalised control (no biomarker present). Significance was determined by ANOVA (*p<0.05, **p<0.01, ***p<0.001, ****p<0.0001, N=3, n=1253-8197).

MULTIPLEXED QUANTIFICATION OF 40 miRNAs

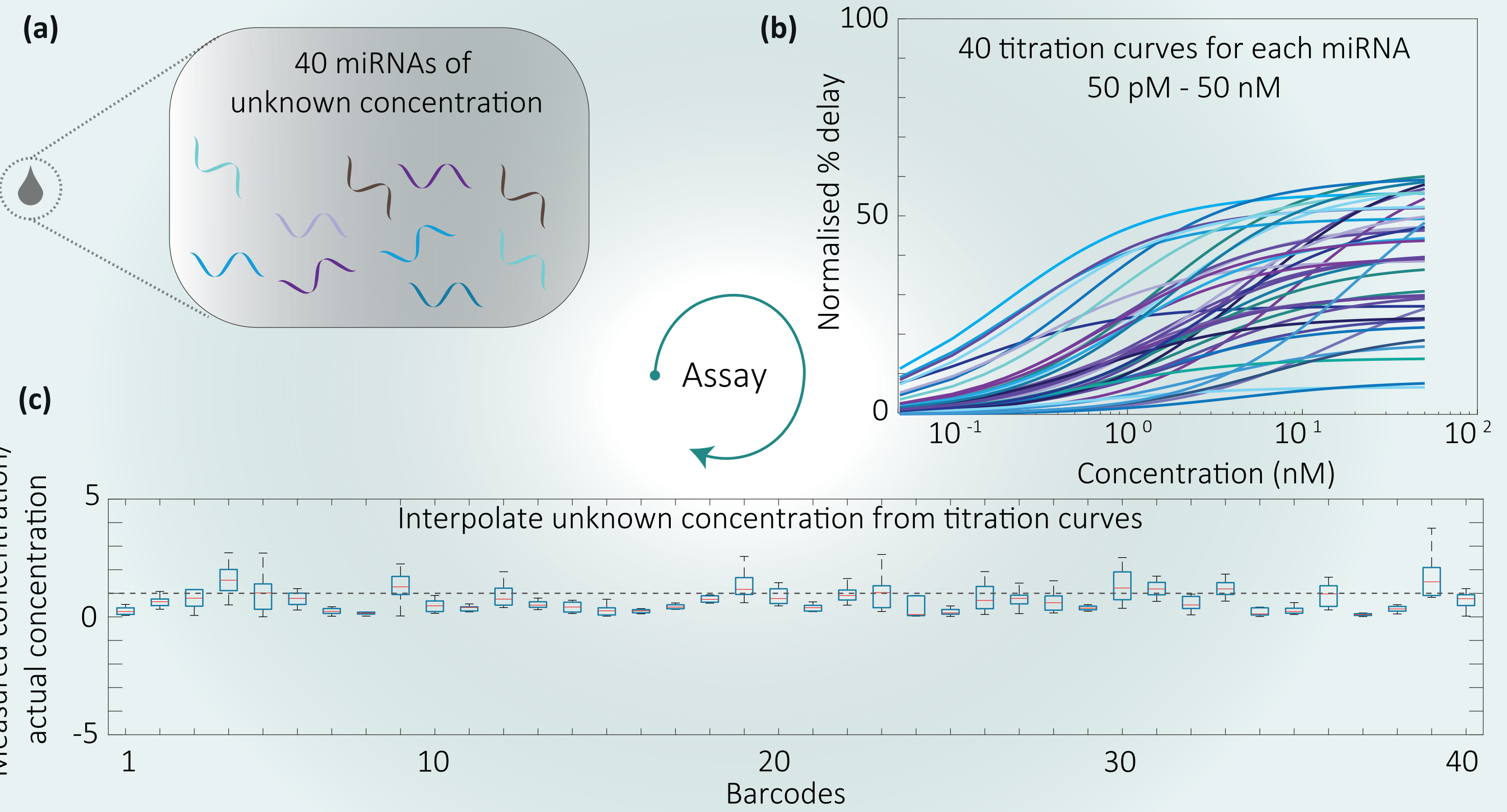


Fig. 3: Quantification of 40 miRNAs. (a) **Unknown concentration of 40 miRNAs** was quantified using (b) **Concentration-/percentage delay curves** of 40 individual barcoded molecular probes derived from a multiplexed experiment (N=5, n=26798-308954). (c) **Single-blinded prediction** of miRNA concentration showing the value for (precited/actual concentration) (N=4, n=6189-71031).

CONCLUSION AND FUTURE WORK

- Individual detection of 40 miRNAs, 4 proteins and 1 small molecule
- Multiplexed detection of miR-208-5p, thrombin and serotonin
- Multiplexed detection and quantification of an unknown concentration of 40 miRNAs
- Detection of miRNAs directly in serum
- Quantification of miRNAs in serum
- Detection of proteins and small molecules in serum
- Quantification of proteins and small molecules
- Transition to kit14 chemistry (ONT) to improve sensitivity
- Expansion of biomarker targets

CLINICAL SAMPLES

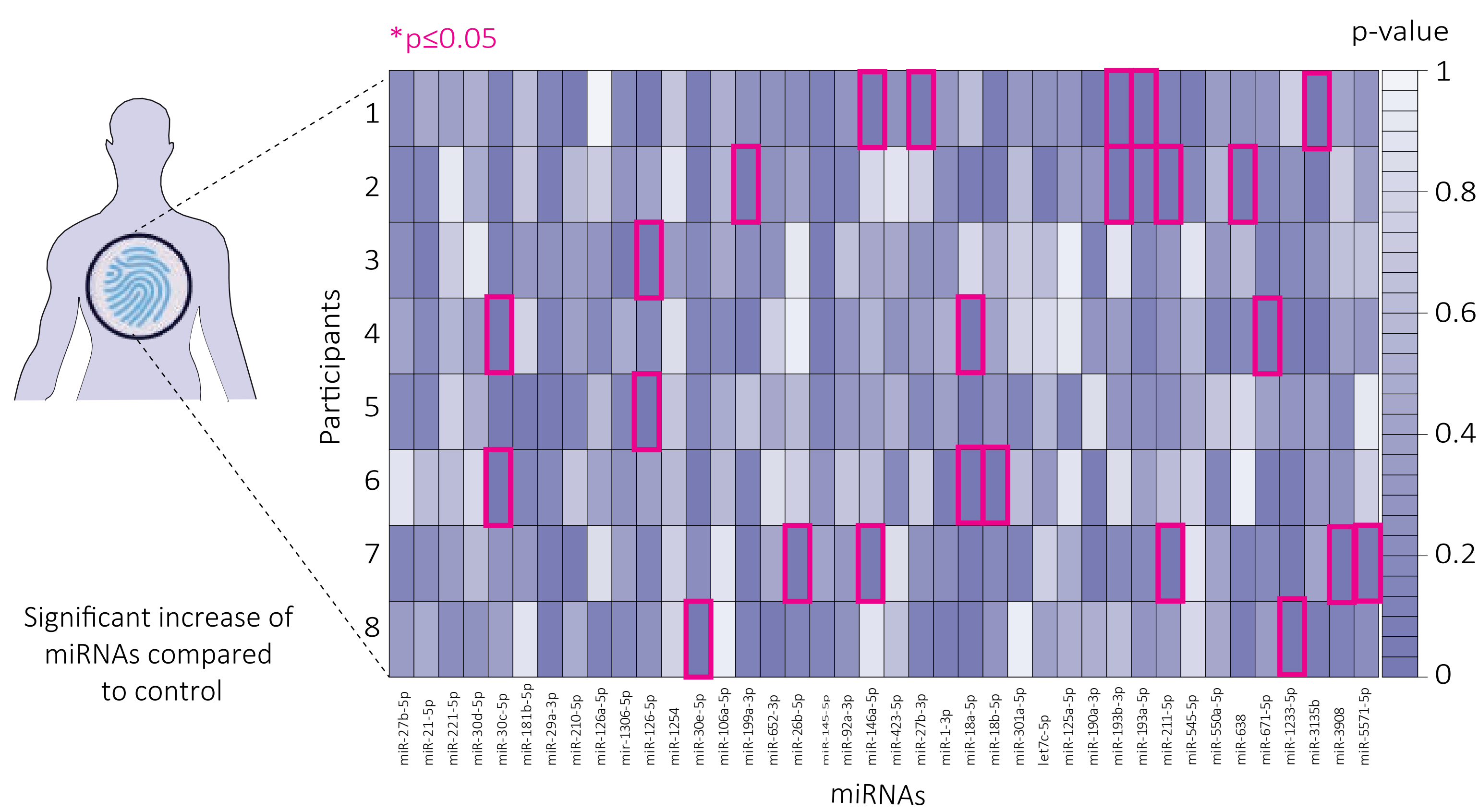


Fig. 4: Detection of 40 miRNAs directly in serum. Heatmap of p-values comparing percentage delayed events +/- serum (Student's t-test, N=3, n=17930-25776).

ACKNOWLEDGEMENT AND REFERENCES

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1 Kasianowicz JJ, Brandin E, Branton D, Deamer DW. (1996) Characterization of individual polynucleotide molecules using a membrane channel. *Proc Natl Acad Sci U S A*.
2 Deamer D, Akeson M, Branton D. (2016) Three decades of nanopore sequencing. *Nat Biotechnol*.
3 Koch C. *et al.* (2022) Highly multiplexed detection of microRNAs, proteins and small molecules using barcoded molecular probes and nanopore sequencing. *Nat Nanotechnol*. (under review)
4 Heron, A, Gutierrez, R, Edel, J, Ivanov, A, Koch, C, Xue, L, Reilly-O'Donnell B. (2022) "Multiplex methods of detecting molecules using nanopores", PCT/GB2022/051268, WO2022243691A1

